

Risk factors for sporadic toxoplasmosis: A systematic review and meta-analysis

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Risk factors for sporadic toxoplasmosis: a systematic review and meta-

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5 **Short Title:** Meta-analysis on risk factors for sporadic toxoplasmosis

6 Abstract

- 7 Toxoplasmosis is considered as the most prevalent parasitic zoonotic infection worldwide.
- 8 The parasitic cycle is mostly known, but the relative contribution of different sources and
- 9 pathways of transmission was not previously studied by a meta-analysis. A systematic review
- and a meta-analysis of case-control, cohort, and cross-sectional studies were performed to
- determine the main risk factors associated with sporadic *Toxoplasma* infection. Suitable
- scientific articles were identified through a systematic literature search and subjected to a
- methodological quality assessment. Mixed-effects meta-analysis models were adjusted by
- population type children, mixed population, and pregnant women to appropriate data
- partitions. 187 primary studies passed the quality assessment stage, investigating risk factors
- 16 for sporadic infection with *Toxoplasma gondii* conducted between 1983 and 2016. Cases were
- 17 defined by serology.

19 The meta-analysis of *Toxoplasma* sporadic infections revealed the significance of

transmission by environmental factors such as contact with soil and contact with animals, in

particular cats. The consumption of raw or undercooked meat and unwashed vegetables

significantly increased the odds of acquiring the disease. Shellfish and raw milk were

identified as significant sources of toxoplasmosis. Almost all meat categories were identified

as risk factors: pork, poultry, beef, processed meat, lamb, and game meat. Contaminated

drinking water may play a role in the acquisition of infection. Moreover, the lack of hygiene

in preparing food was identified as a risk factor. A significant risk factor for pregnant women

is traveling abroad. Lastly, blood transfusion (in pregnant women) and immunocompromised

conditions were found associated with positive serology. The broad definition of exposures

and the use of serology for the case definition are the main limitations for the interpretation of

the results of this meta-analysis. The transmission pathways require further investigations

31 using longitudinal studies and subtyping approaches.

1. Introduction

Toxoplasma gondii, an obligate protozoan parasite of the Apicomplexa phylum, is a worldwide parasite that can infect humans and a large range of warm-blooded vertebrates. Three major clonal lineages (type I to III) differ in pathogenicity and prevalence around the world, with genotype II dominating in congenital toxoplasmosis cases in Europe and the USA (Hosseini et al., 2019). The disease is generally benign, but some severe or life-threatening effects can occur in children (Dunn et al., 1999), when the transmission is congenital, and in immunocompromised patients (Robert-Gangneux and Darde, 2012). Since the conventional designation assumed this clonal population structure, other genotypes have been identified worldwide and termed as "atypical" or "exotic" (Dardé et al., 2014). Among these, highly virulent strains circulating mainly in South America have been responsible for severe cases in immunocompetent people (Carme et al., 2009). Approximately 30% of the human population is considered infected (Montoya and Liesenfeld, 2004). Serological tests are usually used to detect the infection, with detection of anti-T gondii specific IgG and/or IgM antibodies (Montoya, 2002). T. gondii is globally distributed and results in a high public health impact. The Global Disease Burden 2015 Study estimated that foodborne toxoplasmosis was responsible for 10.3 million (95% UI 7.40–14.9 million) cases in 2010, and 825,000 DALYs (95% UI 561,000–1.26 million) DALYs (Torgerson et al., 2015).

The parasitic cycle of toxoplasmosis is complex. During its primary infection, the cat (or other felines), the definitive host, excretes parasites (oocyst form) in its stool. Excretion in cats is limited in time (about two to three weeks) until immunity is established. Oocysts can contaminate the environment: the soil, water, and therefore shellfish that filter water and plant products directly or via irrigation water. Excreted oocysts are not infectious and become infective after sporulation, after few days in the environment depending on climate conditions, and become infectious with long resistance in environmental conditions (oocysts can survive for long periods, up to years, in a favorable environment). The remarkable resistance of the oocyst wall enables the dissemination of *T. gondii* through watersheds and ecosystems, and long-term persistence in diverse foods such as shellfish and fresh produce (Shapiro *et al.*, 2019). Humans and all warm-blooded mammals are infected through the environment or food. Parasites encyst in all tissues, especially striated muscles and the brain. These cysts persist throughout life and can be a source of contamination of new hosts through meat ingestion (carnivorism) (Tenter *et al.*, 2000).

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T. gondii exposure to humans may have multiple origins, and the prevalence is high (and protective for pregnant women). So, numerous epidemiological studies investigate the main transmission pathways of sporadic T. gondii infection by serological studies. A systematic review of outbreaks was recently published (Meireles et al., 2015), still a systematic review and a meta-analysis of case-control, cohort, and cross-sectional studies have to be performed to determine the main risk factors associated with sporadic T. gondii infection. Characterization of risk factors of T. gondii could contribute to identify recommendations for susceptible populations such as pregnant women or immunocompromised patients. The objective of this meta-analysis is to summarize the evidence on risk factors for sporadic T. gondii infection regardless of the country of origin from relevant scientific information contained in epidemiological case-control/cohort/cross-sectional studies.

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2. Material and methods

The protocol of the systematic review and the meta-analysis model are described in depth in the methodological paper of this issue (Gonzales–Barron *et al.*, 2019).

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2.1 Systematic review

- 86 The literature search was conducted between March 2017 and December 2017 using a
- 87 combination of keywords related to (1) Toxoplasma OR Toxoplasmosis, (2) "case-control"
- OR "risk factor" OR cohort (3) infection OR disease, joined by the logical connector AND.
- 89 Relevant studies were identified from five bibliographic search engines, Science Direct,
- 90 PubMed, Scielo, ISI Web of Science and Scopus. No restrictions were defined for the year of
- 91 the study or type of publication. The search was limited to the languages English, French,
- 92 Portuguese and Spanish (Gonzales–Barron et al., 2019).
- 93 Each reference record was screened for relevance for inclusion in the meta-analysis study, and
- 94 subsequently, the methodological quality of the "candidate" studies was assessed using pre-
- 95 set quality criteria comprising (1) appropriate selection of the controls; (2) adjustment to
- 96 correct for confounders, (3) comparability between cases and controls, (4) acceptable
- 97 responses rates for the exposed and control groups; (5) Data analysis appropriate to the study
- 98 design; (6) provision of Odd ratio (OR) with confidence interval or p-value; or provision of
- 99 sufficient data to calculate ORs; overall quality of the study (Gonzales-Barron et al., 2019).
- Primary studies that passed the screening for relevance were marked as having a potential for
- bias if they failed to meet at least one of the methodological quality assessment criteria.

Data from primary studies were extracted using a standardized spreadsheet. Data extracted included the relevant study characteristics (location, time period, population, case definition, design, sample size of the groups, type of model, etc.), the categorized risk factors, the setting, the handling practices and the outcome of the study, odds-ratios (ORs). A data categorization scheme was established to hierarchically group the risk factors into travel, host-specific factors, and pathways of exposure (i.e., person-to-person, animal, environment and, food routes – refer to Gonzales-Barron *et al.*, 2019). In addition to the standard risk factors, the class "Hygiene" (e.g. "no handwashing after toilet", "poor hygiene habits") was also used. Host-specific factors includes blood transfusion.

Considering the parasitic cycle of *T. gondii*, specific partitions were made to investigate more deeply risk factors. The food class "Other red meats" were stratified into lamb, mutton, goat, boar, and venison/horse meat. The route "contact with cats" was also added within "contact with pets". Sexual transmission was found irrelevant and was excluded from the analysis, as humans are not a reservoir of *Toxoplasma*, nor involved in fecal-oral transmission. The variable "Population" was stratified into mixed (adults, or age not defined), pregnant women, and children (under 16 years old). Susceptible population (HIV infection, AIDS, new-borns, liver diseases, chronic kidney diseases, solid organ transplants, mental illnesses, and neurologic diseases) was excluded at the beginning of the study. The acquisition of *Toxoplasma* could be far before the illness (congenital for new-borns) and immunosuppression is by example, a well-known factor of reactivation of a pre-existing infection. Therefore, it was not relevant to study risk factors of acquiring *Toxoplasma* on such populations. Host-specific factors were reduced to immunosuppressive factors that could be linked to a causal pathway of reactivation. Other factors were found irrelevant and excluded because those factors cannot explain the acquisition of the infection (*i.e.* mental illnesses).

2.2 Data synthesis

The full meta-analytical data was first described using basic statistics. Next, data was partitioned into subsets of categories of risk factors ("data partitions"), such as travel, host-specific factors including transmission pathways related to blood transfusion, animal contact, environmental exposures, and food vehicles. Meta-analysis models were then fitted to each of the data partitions, with subgroup class that depends of data partition. For instance, if the data partition was travel, the meta-regression was taking into account the subgroup classes: inside,

abroad, any (Gonzales-Barron et al., 2019). The meta-analytical models were fitted separately 136 137 by population type. For some food classes, the effects of handling (i.e., eating raw, 138 undercooked) and setting (i.e., eating out) on the overall OR were assessed by the calculation 139 of the ratio of the mean OR when food is mishandled to the base OR (Gonzales-Barron et al., 140 2019).

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The statistical analysis was designed to assess the effect of the geographical region, the study period, and the analysis type (univariate/multivariate) on the final result. The objective of the region-specific meta-analysis was to inform the decision on whether the geographical regions were to be maintained for the subsequent pooling of ORs. Geographical regions (Asia, North America, South America, Africa, Europe, Oceania) were removed from a particular metaanalysis partition when their ORs, for this partition, were different from those associated with the other regions or if only less than 3 ORs represented the region (Gonzales-Barron et al., 2019).

All meta-analysis models were substantially weighted random-effects linear regression models. Once a meta-analysis model was fitted, influential diagnostics statistics were applied to remove any influential observation originating from studies marked as having potential-forbias. Publication bias was assessed by funnel plots and statistical tests (Gonzales-Barron et al., 2019) and a statistical test investigating the effect of the study sample size on ORs (Tables 2, 3 and 4) (Gonzalez-Barron et al., 2019). Heterogeneity between studies was assessed by different indicators such as the between-study variability (τ^2), the QE test investigating residual heterogeneity, the variance of residuals, and the intra-class correlation I² (Gonzales-Barron et al., 2019).

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159 All analyses were produced in the R software (R Development Core Team, 2008) implemented with the *metafor* package (Viechtbauer, 2010).

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3. Results

3.1 Descriptive statistics

164 The quality assessment stage was passed by 213 primary studies investigating risk factors for 165 sporadic infection with T. gondii, which were conducted between 1983 and 2016 (80.5% after 166 2000). Excluding susceptible populations other than pregnant women, and some risk factors 167 (see above), 187 publications were retained for meta-analysis (Figure 1 and Appendix 1). 168 Primary studies investigated risk factors in different types of population, namely children (16 169 studies), mixed population (98 studies), and pregnant women (76 studies) (some of them also studied other populations) (Appendix 2). The majority of publications concerns, in descending order, South America (32.5%), Asia (30%), Africa (17.5%), Europe (15%), North America (4%), and Oceania (1%). All publications produced 2050 ORs.

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Toxoplasmosis is generally asymptomatic, so only the presence of antibodies indicates past infection. Symptomatic forms are observed mainly in children infected by congenital transmission (infection occurring during pregnancy in mothers), in immunocompromised people by reactivation of their infection, and more rarely in immunocompetent people. In primary studies, the toxoplasmosis infection was diagnosed by routine antibody screening for T. gondii IgG and/or IgM antibodies. The target populations considered are the mixed population (930 OR), pregnant women (841 OR), and children (185 OR). During the methodological quality assessment, fourteen studies were marked as being possibly affected by bias (Appendix 2). Potential for selection bias was assigned to thirteen studies whose population groups were believed to have a stronger exposure to *Toxoplasma gondii* infection, such as waste pickers and waste workers (Alvarado-Esquivel et al., 2008; Alvarado-Esquivel et al., 2010), livestock and abattoir workers (Adeyisum et al., 2011; Alvarado-Esquivel et al., 2011; Alvarado-Esquivel et al., 2014b), agricultural workers or rural people living in poverty (Alvarado-Esquivel et al., 2013; Cavalcante et al., 2006; Doni et al., 2015; Rostami et al., 2016), veterinary practitioners (Brandon-Mong et al., 2015), inmates (Alvarado-Esquivel et al., 2014a; Sari et al., 2015), and female patients with miscarriages (Tammam et al., 2013). The rationale for assigning a potential-for-bias status to the association measures extracted from Gyang et al. (2015) related to the statistical approach employed, which was a logbinomial model producing prevalence ratio estimates. As explained in Gonzales-Barron et al. (2019), potentially-biased individual ORs were removed only if their influence on the pooled OR estimates was significant, as assessed by the Cook's distance. The risk factors studied concerned the following routes of exposure: host-specific factors (immunocompromised conditions and blood transfusion) (71 OR), lack of personal hygiene (10 OR), travel (21 OR), environmental transmission (476 OR), contact with animals (pets including cats, farm animals, wildlife (511 OR), and food (867 OR).

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3.2 Meta-analysis results

For every data partition, the meta-analyzed risk factors are presented in summary tables only when significant (Tables 1, 2 and 3). Pooled ORs were considered as significant when the lower bound of the 95% CI was equal or greater than 1.Non-significant results for main risk

factors are given in Appendix 3. According to this meta-analysis, travel abroad is a significant risk factor for acquiring positive *Toxoplasma* serology for pregnant women (pooled OR=1.878; 95% CI: 1.284 - 2.746) (Table 1). However, it was non-significant for mixed population, whatever the countries of origin or destination.

Immunocompromised conditions, such as cancer, immunosuppressive treatment or HIV infection, were found to be associated with positive *Toxoplasma* serology in the mixed population (14 publications, pooled OR=2.407; 95% CI: 1.483- 3.909). However, it was non-significant for the pregnant population, with seven publications. Blood transfusion was a significant risk factor for acquiring positive *Toxoplasma* serology in pregnant women (pooled OR=1.785; 95% CI; 1.031 - 3.089) (Forest plot in Figure 2). For the mixed population, blood transfusion did not reach significance (OR=1.368; 95% CI: 0.793 - 2.358). Poor personal hygiene (i.e. "poor hands hygiene") was jointly analyzed for mixed and children populations, and was found significantly associated with positive *Toxoplasma* serology (pooled OR=2.023; 95% CI: 1.693 - 2.416).

Regarding the animal contact pathways, significant associations were found for contact with animals in the mixed, pregnant, and children populations. Occupational contact (raising animals, or contact with animal products) were significant risk factors in both mixed (pooled OR=2.035) and pregnant populations (pooled OR=1.557). Similarly, contact with farm animals was also found significant in the mixed population (pooled OR=1.482; 95% CI: 1.099 - 1.998). Contact with (wild animals) flies/rodents was found significant for all three subpopulations. However, it is a heterogeneous category rather reflective of a relative lack of hygienic conditions of living, with rats or flies inside or near the house (pooled OR from 1.435 to 1.534) (Figure 3). For all the populations studied (mixed, pregnant, or children population), contact with a pet (Table 1), and in particular with cats (Table 2), was a significant risk factor with pooled ORs between 1.631 and 1.711 (Table 2).

For the environmental pathways, consumption of untreated drinking water, farm environment ("living on farm"), contact with wastewater (including "lack of toilets") and contact with soil (such as gardening, playground) were significantly associated with positive *Toxoplasma* serology for the three target populations (except in children population for playground) (Table 1). For the mixed population, 22 ORs coming from Africa region were excluded, due to the high level of OR, concerning untreated drinking water (9), farm environment (6), and

- 238 playground (7). Of course, considering them is not changing the significativity of results
- concerning those relevant categories (results not shown).
- For the children population, one OR from an Oceanian study was excluded, because it was
- isolated. Six ORs coming from Africa were also excluded, describing exposure to drinking
- unboiled water and playground, because of the low values of their OR, in comparison with
- other regions. For the children population, the contact with soil and garden ("playground")
- was close to reaching significance (pooled OR=1.138; 95% CI: 0.969 1.335).

- 246 The meta-analysis on food consumption pathways revealed significant associations with meat
- for the mixed population (pooled OR=1.761; 95% CI: 1.570 1.974) and pregnant women
- 248 (pooled OR=1.960; 95% CI: 1.472 2.610) (Table 1). By contrast, in the children population
- 249 from six publications from South America and Africa, the overall OR associated with meat
- was not significant (pooled OR=1.329; 95% CI: 0.891 1.985; Appendix 3). For the mixed
- 251 population, almost all meat matrices were identified as risk factors: pork meat (pooled OR=
- 252 2.114; 95% CI: 1.411 3.169), poultry (pooled OR=1.623; 95% CI: 1.147 2.297), beef
- 253 (pooled OR=1.635; 95% CI: 1.170 2.285), processed meat like salami or sausages (pooled
- 254 OR=1.365; 95% CI: 1.075 1.733), other red meat (goat, mutton or lamb) (pooled OR=
- 255 1.897; 95% CI: 1.480 2.431) and other types of meat (game or undercooked meat) (pooled
- 256 OR=1.734; 95% CI: 1.544 1.947]) (Table 2). For this population, when it was possible, more
- precise items could be analyzed such as wild boar meat (pooled OR=2.487; 95% CI: 1.814 -
- 258 3.409), lamb meat (pooled OR=2.404; 95% CI: 1.189 4.859), goat meat (pooled OR=1.667;
- 259 95% CI: 1.212 2.294), and other types of red meat (such as venison, ram and horse meat)
- 260 (pooled OR=1.566; 95% CI: 1.221 2.008) (Table 2).

- In pregnant women, significant sources were beef (pooled OR=2.052; 95% CI: 1.576 2.672),
- other red meat (goat, mutton or lamb) (pooled OR=1.822; 95% CI: 1.279 2.595), poultry
- 264 (pooled OR=1.514; 95% CI: 1.130 2.028), processed meat, like cured meat, sausages and
- salami (pooled OR=1.532; 95% CI: 1.201 1.953), and other meat such as undercooked or
- raw meat (most often in this category) and game meat (sometimes) (pooled OR=1.592; 95%
- 267 CI: 1.354 1.871) (Table 2). The pork consumption was not found significant for pregnant
- women with a pooled OR=1.04 (95% CI: 0.780 1.387). Within the red meat category
- 269 (including lamb and mutton/sheep), only the consumption of lamb was found significant
- 270 (pooled OR=1.832; 95% CI: 1.148 2.922; Table 2). In children, meat, dairy products, and

produce consumption were not found significantly associated with positive *Toxoplasma* serology.

The consumption of produce was associated with positive *Toxoplasma* serology for the mixed population (pooled OR=1.872; 95% CI: 1.539 - 2.276) and pregnant women (pooled OR=1.651; 95% CI: 1.267 - 2.151) (Table 1). Within produce, only vegetables were identified as a risk factor for the mixed population (pooled OR=1.866; 95% CI: 1.491 - 2.335) and pregnant women (pooled OR=1.372; 95% CI: 1.198 - 1.571) (Table 2). The consumption of seafood in the mixed population (not studied in pregnant nor in children population) (Table1), in particular shellfish ("mollusks") (Table 2), was found in association with positive *Toxoplasma* serology (pooled OR=1.917; 95% CI: 1.395 - 2.636) (Figure 5). Nevertheless, the fish consumption was not found significant associated with toxoplasmosis.

The consumption of eggs was not found to be a determinant of positive *Toxoplasma* serology either in the mixed or in the pregnant population. Dairy products (mainly raw milk) were significant risk factors for the mixed population (pooled OR=1.563; 95% CI: 1.298 - 1.882) (Figure 4) and pregnant women (pooled OR=1.521; 95% CI: 1.116 - 2.073) (Table 1). The consumption of raw milk or raw milk cheese in comparison with pasteurized dairy products significantly increases the risk by a factor of 1.430 (95% CI: 1.073 - 1.905; Table 3). Regarding the effect of the food practices (Table 3), eating undercooked or raw meats or unwashed produce were identified as well as risk factors. The poor handling of foods (mainly "not washing hands before eating" or "using unwashed kitchen utensils") was also found associated with positive *Toxoplasma* serology in pregnant women (Table 1).

For most of the meta-analytical models reported in Tables 1, 2, and 3 the statistical tests indicated the absence of potential significant publication bias. Exceptions were observed in partitions related to food (Table 1), meat (Table 2), for mixed and pregnant population, and practices of cooking meat (Table 3) (all populations). The funnel plots for food and meat in the mixed population (Figure 6) evidences asymmetry at its basis, in an overestimation manner, and underestimation at the top. The weight given in meta-analysis is lower in study outcomes located at the basis than in those falling at the top of the funnel. The funnel plots for food and meat in pregnant women, as for meat cooking practices, show little asymmetry that may lead to some overestimation (Figure 6). The intra-class correlation I² indicates significant low (<25%) to moderate (<50% or around 50%) remaining between-study heterogeneity in all

the data partitions. As Q or QE statistic was still significant, the moderator(s) already considered in the meta-analysis models could not fully explain the between-study heterogeneity (Gonzales-Barron et al., 2019).

4. Discussion

The measurement of seroprevalence is an indicator of *T. gondii* infection, but it does not provide information on the time of infection. Besides, there may be a significant lapse of time between contamination and the collection of information on the exposures of infected persons (most often identified by serological research of IgG) during studies. Consequently, the results are conditioned on the absence of any change in the respondents' exposures over time. Cases could have been identified through serological tests whose performances (sensitivity and/or specificity) are not necessarily equivalent between countries and over time. However, we assume that these meta-analyses are not particularly biased (over or under-estimation) due to those parameters. Furthermore, the origin of the infection (congenital or postnatally-acquired) is not known in the primary studies included in this meta-analysis (case-control or transversal studies). Nevertheless, the relative share of congenital infections in the measurement of seroprevalence in children and adults is supposed to be low.

Considering the high prevalence of *Toxoplasma* infection globally, and the diversity of pathways and sources of exposure to humans, the number of publications and investigated risk factors is large and, most publications are recent. In comparison with the systematic reviews of other foodborne pathogens of this special issue, this is the one with the highest number of included case-control, cohort, and cross-sectional studies. As toxoplasmosis outbreaks are rare, studying sporadic cases is the main way to identify potential sources, and to make specific recommendations to populations at-risk such as pregnant women or immunocompromised individuals. Then, our results are based most often on a considerable number of outcomes and are in agreement with other published evidence.

Blood transfusion was identified as a source of toxoplasmosis in this meta-analysis, which may seem at first surprising, but it has been reported as a risk factor in recent case-control studies and molecular tools allow the detection of *Toxoplasma* DNA in blood samples (Saki *et al.*, 2019). Blood transfusion is thought to be a rare source of transmission because the duration of parasitemia during acute infection is brief but transmission by thachyzoites stage is conceivable. Immunocompromised conditions were found to be associated with

Toxoplasma gondii infection in mixed population: this result is in agreement with a recent meta-analysis (Wang et al., 2017). However, its interpretation is not straightforward because the serological methods could be less sensitive and specific in immunocompromised individuals (Wang et al., 2017).

As expected, exposure to cats appeared as a significant risk factor, which has long been recognized in many case-control studies (Cook *et al.*, 2000). Felids, specifically the domestic cat, have a central role as the source of infective oocysts; the implications of increasing cat populations as a public health risk due to toxoplasmosis have been reviewed in Dabritz and Conrad (2010). One consequence has been increased *T. gondii* oocyst dissemination to the environment, associated with a higher risk of transmission to humans and animals. Significant risk factors as "contact with other animals" are probably due to unhygienic contamination, concerning environmental contamination (e.g. soil on the fur), and "contact with flies or rodents" in children, could reflect indirect contamination. Although this risk factor is not often studied, the role of flies and other pests has been identified as vectors by oocyst transport (Frenkel *et al.* 1995). Environmental transmission by oocyst stage appears to be influential; identified risk factors are contact with wastewater associated with poor hygiene, contact with soil, farm environment that aggregates different risk factors not clearly defined, and consumption of untreated or inadequately treated water.

For occupational exposure, farm animals exposure, and farm environment, more precise definition about the raised animals, e.g., sheep vs. cattle vs. pig farming vs. mixed, or their raising conditions, in future studies, could allow to break down those categories for further meta-analysis investigation. Our results are in agreement with a systematic review of outbreaks that showed that in 38 outbreaks (worldwide), 21% could be attributed to contaminated waters and 26% to contact with contaminated soil (Meireles *et al.*,2015). Contamination of water and soils by oocysts was shown in different studies (Aubert and Villena, 2009; Bahia-Oliveira, 2017; Bahia-Oliveira *et al.*, 2003; Shapiro *et al.*, 2019).

With regards to food consumption, undercooked meat, unwashed vegetables, raw milk, and shellfish are risk factors for positive *Toxoplasma* serology. Those results are in accordance with the analysis of published outbreaks, showing that raw or undercooked meat was the origin of 44.7% of the outbreaks and raw vegetables of 5.3% (Meireles *et al.*, 2015). Furthermore, our results are in line with the meta-analysis conducted by Belluco *et al.* (2018) to assess the risk of humans developing acute *T. gondii* infection, due to the consumption of

undercooked beef, sheep (or meat), which were also found significant. In their study, consumption of raw/undercooked pork, raw eggs, and unpasteurized milk were non-significant risk factors. Concerning the role of meat, raw or undercooked meats were identified as significant risk factors for toxoplasmosis. Within meats, with except for wild boar, the type of meat found is consistent with the literature. Five publications, all conducted in Mexico, identified wild boar meat as a risk factor (which is a low number of studies compared to other risk factors). Game meat also stands out significantly. *T. gondii* has been detected in sheep and lamb (Dubey, 2009; Halos *et al.*, 2010), cattle (Dubey 1986;1992; Fromont *et al.*, 2009; Blaga *et al.*, 2019), pork (Djokic *et al.*, 2016; Foroutan *et al.*, 2019), wild boar (Roqueplo *et al.*, 2017; Rostami *et al.*, 2017) and also other animals like poultry (Tonouhewa, 2017; Guo *et al.*, 2016).

The identification of raw milk as a risk factor is based on 16 publications for the mixed population and 27 for pregnant women. A recent review on milk consumption and *Toxoplasma* infection reported mainly goat milk as a source of infection (Boughattas, 2017). Moreover, *T. gondii* has been detected in raw milk from infected animals (Cisak *et al.*, 2017; Vismarra *et al.*, 2017). Further studies should investigate the contamination in milk (sheep/cows), and the spatial and temporal variability of contamination in farms and animals could be better explored in the future. Shellfish (oysters, mussels, etc.) are identified from four studies conducted in Taiwan (2), the United States (1), and the United Kingdom (1) only. Shellfish contamination has been recently established (Cong *et al.*, 2017; Ghozzi *et al.*, 2017; Coupe *et al.*, 2018). However, compared with meat-producing and poultry animals, research on seafood species contaminated with *T. gondii* represents a relatively new field of study, and harmonized methods of oocysts detection are lacking (Shapiro *et al.*, 2019).

Lack of hygiene, in particular, the lack of handwashing before eating or preparing a meal, was also found to play a role in the acquisition of *Toxoplasma* infection. Produce, in particular unwashed vegetables, was a significant risk factor which is expected considering the life cycle of the parasite. The contamination of vegetables was explored in few studies, but data are lacking in numerous countries due to the absence of standard detection methods for *T. gondii* oocysts in these matrices (Shapiro *et al.*, 2019). The vast range of types of vegetables, not described in the studies, could be better investigated (i.e., leaf or root vegetables).

5. Conclusion

The risk factors identified in this meta-analysis could complement those already established in future sporadic case-control studies quantifying the attributable risk fraction: mollusks (regarding the species), raw milk (goat/cows), vegetables (including the type of vegetable and the preparation i.e., washing), game meat and drinking water. Furthermore, the development of sensitive methods for the detection and isolation of *T. gondii* in these matrices is needed to confirm the causal association revealed in epidemiological studies. In the context of the global market of food and animals, it is essential to obtain information from an international meta-analysis. This meta-analysis can be seen as a tool to identify emerging situations. The transmission pathways need further investigations using longitudinal studies (to avoid memory bias) and subtyping approaches (to differentiate between strain infectivity). Information on the frequency or the duration of exposure could be further investigated. Investigation of sporadic cases of infection remains essential for further understanding of *Toxoplasma gondii* transmission.

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References

- 1. Adesiyun, A., Campbell, M., Rahaman, S., Bissessar, S., Stewart-Johnson, A., Dookeran,
- S., and Hilaire, M.G.-S. (2011). Frequency of detection of immunoglobulins of
- 436 Toxoplasma gondii, Leptospira spp., and Brucella abortus in livestock/farm and abattoir
- workers in Trinidad. *Journal of Agromedicine*, 16(3):200–209
- 438 2. Alvarado-Esquivel, C., Liesenfeld, O., Márquez-Conde, J., Cisneros-Camacho, A.,
- Estrada-Martínez, S., Martínez-García, S., González-Herrera, A., GarcíaCorral, N.,
- 440 (2008). Seroepidemiology of infection with *Toxoplasma gondii* in waste pickers and waste
- workers in Durango, Mexico. *Zoonoses and Public Health*, 55(6):306–312.
- 3. Alvarado-Esquivel, C., Liesenfeld, O., Márquez-Conde, J., Estrada-Martínez, S., Dubey,
- J., (2010). Seroepidemiology of infection with Toxoplasma gondii in workers
- occupationally exposed to water, sewage, and soil in Durango, Mexico. Journal of
- 445 *Parasitology*, 96(5):847–850.
- 446 4. Alvarado-Esquivel, C., Liesenfeld, O., Estrada-Martínez, S., Félix-Huerta, J. (2011).
- 447 Toxoplasma gondii infection in workers occupationally exposed to raw meat.
- 448 Occupational Medicine-Oxford, 61(4):265–269.
- 5. Alvarado-Esquivel, C., Campillo-Ruiz, F., Liesenfeld, O., (2013). Seroepidemiology of
- infection with Toxoplasma gondii in migrant agricultural workers living in poverty in
- Durango, Mexico. Parasites & vectors, 6, 113.
- 452 6. Alvarado-Esquivel, C., Hernández-Tinoco, J., Sánchez-Anguiano, L.F., Ramos-Nevárez,
- 453 A., Cerrillo-Soto, S.M., Sáenz-Soto, L., Liesenfeld, O., (2014a). High seroprevalence of
- 454 Toxoplasma gondii infection in inmates: A case control study in Durango city, Mexico.
- 455 European Journal of Microbiology & Immunology, 4(1):76–82.
- 456 7. Alvarado-Esquivel, C., Pacheco-Vega, S.J., Hernández-Tinoco, J., Saldaña-Simental,
- D.E., Sánchez-Anguiano, L.F., Salcedo-Jáquez, M., Ramos-Nevárez, A., Liesenfeld, O.,
- Márquez-Conde, J.A., Cerrillo-Soto, S.M., Martínez-Ramírez, L., Guido-Arreola, C.A.,
- 459 (2014b). Lack of association between Toxoplasma gondii infection and occupational
- exposure to animals. European Journal of Microbiology & Immunology, 4(4):184–192.
- 8. Aubert D. and Villena I. (2009). Detection of Toxoplasma gondii oocysts in water:
- proposition of a strategy and evaluation in Champagne-Ardenne Region, France. *Mem Inst*
- 463 *Oswaldo Cruz.*104(2):290-295.

- 9. Bahia-Oliveira, L.M., Jones, J.L., Azevedo-Silva, J., Alves, C.C., Orefice, F., Addiss,
- D.G., (2003). Highly endemic, waterborne toxoplasmosis in north Rio de Janeiro state,
- 466 Brazil. *Emerging Infectious Diseases* 9:55–62.
- 467 10. Bahia-Oliveira, L., Gomez-Marin, J., Shapiro, K., Jiménez-Cisneros, A., (2017).
- Toxoplasma gondii. In: Rose, J.B., B. (Eds.), Global Water Pathogens Project. Michigan
- State University, E. Lansing, MI, UNESCO.
- 470 11. Belluco, S., Simonato, G., Mancin, M., Pietrobelli, M., Ricci, A., (2018). Toxoplasma
- 471 gondii infection and food consumption: A systematic review and meta-analysis of case-
- 472 controlled studies. *Critical reviews in food science and nutrition*, 58(18):3085–3096.
- 473 12. Blaga, R., Aubert, D., Thébault, A., Perret, C., Geers, R., Thomas, M., Alliot, A., Djokic,
- V., Ortis, N., Halos, L., Durand, B., Mercier, A., Villena, I., & Boireau, P. (2019).
- 475 Toxoplasma gondii in beef consumed in France: regional variation in seroprevalence and
- parasite isolation. *Parasite* (Paris, France), 26:77.
- 477 13. Boughattas, S. (2017) Toxoplasma infection and milk consumption: Meta-analysis of
- 478 assumptions and evidences. *Critical reviews in food science and nutrition*, 57, 2924-2933.
- 479 14. Brandon-Mong, G. J., Che Mat Seri, N. A., Sharma, R. S., Andiappan, H., Tan, T. C.,
- Lim, Y. A., Nissapatorn, V., (2015). Seroepidemiology of Toxoplasmosis among People
- Having Close Contact with Animals. *Frontiers in immunology*, 6, 143.
- 482 15. Carme, B., Demar, M., Ajzenberg, D., Dardé, M. L., (2009). Severe acquired
- 483 toxoplasmosis caused by wild cycle of Toxoplasma gondii, French Guiana. Emerging
- 484 infectious diseases, 15(4):656–658.
- 485 16. Cavalcante, G., Aguiar, D., Camargo, L., Labruna, M., Andrade, H., Meireles, L., Dubey,
- J., Thulliez, P., Dias, R., Gennari, S., (2006). Seroprevalence of Toxoplasma gondii
- antibodies in humans from rural Western Amazon, Brazil. Journal of Parasitology,
- 488 92(3):647–649.
- 489 17. Cisak, E., Zając, V., Sroka, J., Sawczyn, A., Kloc, A., Dutkiewicz, J., Wójcik-Fatla, A.,
- 490 (2017). Presence of Pathogenic Rickettsiae and Protozoan in Samples of Raw Milk from
- Cows, Goats, and Sheep. Foodborne pathogens and disease, 14(4):189–194.
- 492 18. Cong, W., Zhang, N. Z., Hou, J. L., Wang, X. C., Ma, J. G., Zhu, X. Q., Chen, G. J.,
- 493 (2017). First detection and genetic characterization of Toxoplasma gondii in market-sold
- 494 oysters in China. Infection, Genetics and Evolution: Journal of Molecular Epidemiology
- 495 and Evolutionary Genetics in Infectious Diseases, 54:276-278.
- 496 19. Cook, A. J., Gilbert, R. E., Buffolano, W., Zufferey, J., Petersen, E., Jenum, P. A., Foulon,
- W., Semprini, A. E., Dunn, D. T., (2000). Sources of toxoplasma infection in pregnant

- women: European multicentre case-control study. European Research Network on
- 499 Congenital Toxoplasmosis. *BMJ (Clinical research ed.)*, 321(7254): 142–147.
- 500 20. Coupe, A., Howe, L., Burrows, E., Sine, A., Pita, A., Velathanthiri, N., Vallée, E.,
- Hayman, D., Shapiro, K., & Roe, W. D. (2018). First report of Toxoplasma gondii
- sporulated oocysts and Giardia duodenalis in commercial green-lipped mussels (Perna
- canaliculus) in New Zealand. *Parasitology research*, 117(5):1453–1463.
- 504 21. Dabritz, H. A. and P. A. Conrad (2010). Cats and Toxoplasma: Implications for Public
- Health. *Zoonoses and Public Health*, 57(1): 34-52.
- 506 22. Dardé, ML, Ajzenberg, D and Su, C (2014). Molecular epidemiology and population
- structure of Toxoplasma gondii. Toxoplasma gondii (Second Edition). The Model
- Apicomplexan-Perspectives and Methods. pp. 61–67.
- 509 23. Djokic, V., R. Blaga, D. Aubert, B. Durand, C. Perret, R. Geers, T. Ducry, I. Vallee, O.
- 510 Djurkovic Djakovic, A. Mzabi, I. Villena, Boireau P., (2016): *Toxoplasma gondii* infection
- in pork produced in France. *Parasitology*, 143:557-567.
- 512 24. Doni, N., Simsek, Z., Gurses, G., Zeyrek, F., and Demir, C., (2015). Prevalence and
- associated risk factors of Toxoplasma gondii in female farmworkers of Southeastern
- Turkey. *Journal of Infection in Developing Countries*, 9(1):087–093.
- 515 25. Dubey JP. (1986). A review of toxoplasmosis in cattle. *Veterinary Parasitology*, 22(3-4):
- 516 177-202.
- 517 26. Dubey JP. (1992). Isolation of *Toxoplasma gondii* from a naturally infected beef cow.
- 518 *Journal of Parasitology*, 78(1):151-153.
- 519 27. Dubey JP. (2009). Toxoplasmosis in sheep. The last 20 years. *Veterinary Parasitology*;
- 520 163:1–14.
- 521 28. Dunn, D., Wallon, M., Peyron, F., Petersen, E., Peckham, C., Gilbert, R. (1999). Mother-
- 522 to-child transmission of toxoplasmosis: risk estimates for clinical counselling. Lancet
- 523 (London, England), 353(9167): 1829–1833.
- 524 29. Foroutan, M., Fakhri, Y., Riahi, S. M., Ebrahimpour, S., Namroodi, S., Taghipour, A.,
- 525 Spotin, A., Gamble, H. R., & Rostami, A. (2019). The global seroprevalence of
- 526 Toxoplasma gondii in pigs: A systematic review and meta-analysis. Veterinary
- 527 *parasitology*, 269: 42–5
- 30. Frenkel, J. K., Hassanein, K. M., Hassanein, R. S., Brown, E., Thulliez, P., & Quintero-
- Nunez, R. (1995). Transmission of Toxoplasma gondii in Panama City, Panama: a five-
- year prospective cohort study of children, cats, rodents, birds, and soil. *The American*
- journal of tropical medicine and hygiene, 53(5), 458–468.

- 31. Fromont, E. G., Riche, B., & Rabilloud, M. (2009). Toxoplasma seroprevalence in a rural
- population in France: detection of a household effect. *BMC infectious diseases*, 9, 76.

- 32. Gilot-Fromont, E., D. Aubert, S. Belkilani, P. Hermitte, O. Gibout, R. Geers and I. Villena
- 536 (2009). Landscape, herd management and within-herd seroprevalence of Toxoplasma
- gondii in beef cattle herds from Champagne-Ardenne, France. *Veterinary Parasitology*
- 538 161(1): 36-40.
- 33. Ghozzi, K., M. Marangi, R. Papini, I. Lahmar, R. Challouf, N. Houas, R. Ben Dhiab, G.
- Normanno, H. Babba, Giangaspero A. (2017). First report of Tunisian coastal water
- contamination by protozoan parasites using mollusk bivalves as biological indicators.
- 542 *Marine Pollution Bulletin.* 117(1): 97-202.
- 34. Gonzales-Barron, U., Thébault, A., Kooh, P., Watier, L., Sanaa, M., Cadavez, V., (2019).
- Strategy for systematic review of observational studies and meta-analysis modelling of
- risk factors for sporadic foodborne diseases. Microbial risk analysis, 10082. doi:
- 546 10.1016/j.mran.2019.07.003
- 35. Guo, M., Mishra, A., Buchanan, R. L., Dubey, J. P., Hill, D. E., Gamble, H. R., Jones, J.
- L., Pradhan, A. K., (2016). A Systematic Meta-Analysis of Toxoplasma gondii Prevalence
- in Food Animals in the United States. *Foodborne pathogens and disease*, 13(3):109–118.
- 36. Gyang, V., Akinwale, O., Lee, Y.-L., Chuang, T.-W., Orok, A., Ajibaye, O., Liao, C.-W.,
- 551 Cheng, P.-C., Chou, C.-M., Huang, Y.-C., Fan, K.-H., C.-K., Fan., (2015). *Toxoplasma*
- 552 gondii infection: Seroprevalence and associated risk factors among primary
- schoolchildren in Lagos city, Southern Nigeria. Revista da Sociedade Brasileira de
- 554 *Medicina Tropical*, 48(1):56–63.
- 555 37. Halos, L., Thébault, A., Aubert, D., Thomas, M., Perret, C., Geers, R., Alliot, A., Escotte-
- Binet, S., Ajzenberg, D., Dardé, M. L., Durand, B., Boireau, P., Villena, I., (2010). An
- innovative survey underlining the significant level of contamination by Toxoplasma
- gondii of ovine meat consumed in France. International journal for parasitology,
- *40*(2):193–200.
- 38. Hosseini, S. A., Amouei, A., Sharif, M., Sarvi, S., Galal, L., Javidnia, J., Pagheh, A. S.,
- Gholami, S., Mizani, A., & Daryani, A. (2018). Human toxoplasmosis: a systematic
- review for genetic diversity of Toxoplasma gondii in clinical samples. *Epidemiology and*
- 563 infection, 147, E36:1–9.

- 564 39. Meireles, L. R., Ekman, C. C., Andrade, H. F., Jr, Luna, E. J. (2015). Human
- toxoplasmosis outbreaks and the agent infecting form. findings from a systematic review.
- *Revista do Instituto de Medicina Tropical de Sao Paulo*, 57(5): 369-376.
- 567 40. Montoya JG. (2002). Laboratory diagnosis of Toxoplasma gondii infection and
- toxoplasmosis. *J Infect Dis.* 2002;185 Suppl 1:S73-S82. doi:10.1086/338827
- 41. Montoya, J. G. and O. Liesenfeld (2004). Toxoplasmosis. *Lancet* 363(9425):1965-1976.
- 570 42. Robert-Gangneux, F. and M. L. Darde (2012). Epidemiology of and diagnostic strategies
- for toxoplasmosis. *Clinical Microbiology Reviews* 25(2):264-296.
- 572 43. Roqueplo, C., Blaga, R., Marié, J., Vallée, I., Davoust, B., (2017). Seroprevalence of
- 573 Toxoplasma gondii in hunted wild boars (Sus scrofa) from southeastern France. Folia
- *Parasitologica*, 64, 2017.000.
- 575 44. Rostami, A., Seyyedtabaei, S., Aghamolaie, S., Behniafar, H., Lasjerdi, Z., Abdolrasouli,
- A., Mehravar, S., Alvarado-Esquivel, C. (2016). Seroprevalence and risk factors
- associated with *Toxoplasma gondii* infection among rural communities in Northern Iran.
- 578 Revista do Instituto de Medicina Tropical de São Paulo, 58, 70.
- 579 45. Rostami, A., Riahi, S. M., Fakhri, Y., Saber, V., Hanifehpour, H., Valizadeh, S.,
- Gholizadeh, M., Pouya, R. H., Gamble, H. R., (2017). The global seroprevalence of
- Toxoplasma gondii among wild boars: A systematic review and meta-analysis. *Veterinary*
- 582 *parasitology*, 244:12–20.
- 583 46. Saki, J., Foroutan, M., Khodkar, I., Khodadadi, A., Nazari, L., (2019). Seroprevalence and
- molecular detection of Toxoplasma gondii in healthy blood donors in southwest Iran.
- *Transfusion and apheresis science*, 58(1):79–82
- 586 47. Sari, Y., Haryati, S., Raharjo, I., Prasetyo, A.A., (2015). *Toxoplasma* and viral antibodies
- among HIV patients and inmates in central Java, Indonesia. Southeast Asian Journal of
- *Tropical Medicine and Public Health*, 46(6):977–985.
- 589 48. Shapiro, K., L. Bahia-Oliveira, B. Dixon, A. Dumètre, L. A. de Wit, E. VanWormer,
- Villena I., (2019). Environmental transmission of Toxoplasma gondii: Oocysts in water,
- soil and food. *Food and Waterborne Parasitology* 15: e00049.
- 592 49. Shapiro, K., Kim, M., Rajal, V. B., Arrowood, M. J., Packham, A., Aguilar, B., & Wuertz,
- 593 S., (2019). Simultaneous detection of four protozoan parasites on leafy greens using a
- novel multiplex PCR assay. *Food microbiology*, 84, 103252.
- 595 50. Tammam, A., Haridy, M., Abdellah, A., Ahmed, S., Fayed, H., Alsammani, M., (2013).
- Seroepidemiology of *Toxoplasma Gondii* infection in women with first trimester

- spontaneous miscarriage in Qena Governorate, Egypt. Journal of Clinical and Diagnostic
- 598 *Research*, 7(12):2870–2873.
- 599 51. Tenter, A. M., Heckeroth, A. R., & Weiss, L. M. (2000). Toxoplasma gondii: from
- animals to humans. *International journal for parasitology*, 30(12-13): 1217–1258.
- 52. Tonouhewa, A. B., Akpo, Y., Sessou, P., Adoligbe, C., Yessinou, E., Hounmanou, Y. G.,
- Assogba, M. N., Youssao, I., Farougou, S., (2017). *Toxoplasma gondii* infection in meat
- animals from Africa: Systematic review and meta-analysis of sero-epidemiological
- 604 studies. *Veterinary world*, 10(2):194–208.
- 53. Torgerson, P. R., Devleesschauwer, B., Praet, N., Speybroeck, N., Willingham, A. L.,
- Kasuga, F., Rokni, M. B., Zhou, X. N., Fèvre, E. M., Sripa, B., Gargouri, N., Fürst, T.,
- Budke, C. M., Carabin, H., Kirk, M. D., Angulo, F. J., Havelaar, A., de Silva, N., (2015).
- World Health Organization Estimates of the Global and Regional Disease Burden of 11
- Foodborne Parasitic Diseases, 2010: A Data Synthesis. *PLoS medicine*, 12(12), e1001920.
- 610 https://doi.org/10.1371/journal.pmed.1001920
- 54. Vismarra, A., Barilli, E., Miceli, M., Mangia, C., Bacci, C., Brindani, F., Kramer, L.,
- 612 (2017). Toxoplasma Gondii and Pre-treatment Protocols for Polymerase Chain Reaction
- Analysis of Milk Samples: A Field Trial in Sheep from Southern Italy. *Italian journal of*
- 614 *food safety*, 6(1):6501

619

- 615 55. Wang, Z. D., Liu, H. H., Ma, Z. X., Ma, H. Y., Li, Z. Y., Yang, Z. B., Zhu, X. Q., Xu, B.,
- Wei, F., Liu, Q., (2017). *Toxoplasma gondii* Infection in Immunocompromised Patients:
- A Systematic Review and Meta-Analysis. Frontiers in microbiology, 8, 389.

621 FIGURE CAPTIONS 622 Figure 1: Flow chart of literature search for case-control, cohort and cross-sectional studies of 623 human sporadic toxoplasmosis 624 625 Figure 2: Forest plot of the association of blood transfusion with positive *Toxoplasma* 626 serology in pregnant women 627 628 Figure 3: Forest plot of the association of flies/rodents with positive *Toxoplasma* serology in 629 the mixed population 630 631 Figure 4: Forest plot of the association of consumption of dairy products with positive 632 Toxoplasma serology in the mixed population 633 Figure 5: Forest plot of the association of consumption of molluscs with positive *Toxoplasma* 634 635 serology in the mixed population 636 637 Figure 6: Funnel plots of meta-analyses investigating categorized risk factors (i.e., food in 638 mixed and pregnant population, meat in mixed and pregnant population, and practices of 639 cooking meat) 640 641 LIST OF TABLES 642 Table 1. Meta-analysis results on main (significant) risk factors for positive *Toxoplasma* 643 serology 644 Table 2. Meta-analysis results on disaggregated risk factors for positive *Toxoplasma* serology 645 Table 3. Effect of poor handling on the pooled association between positive Toxoplasma 646 serology and consumption of meat, vegetables and dairy 647 648

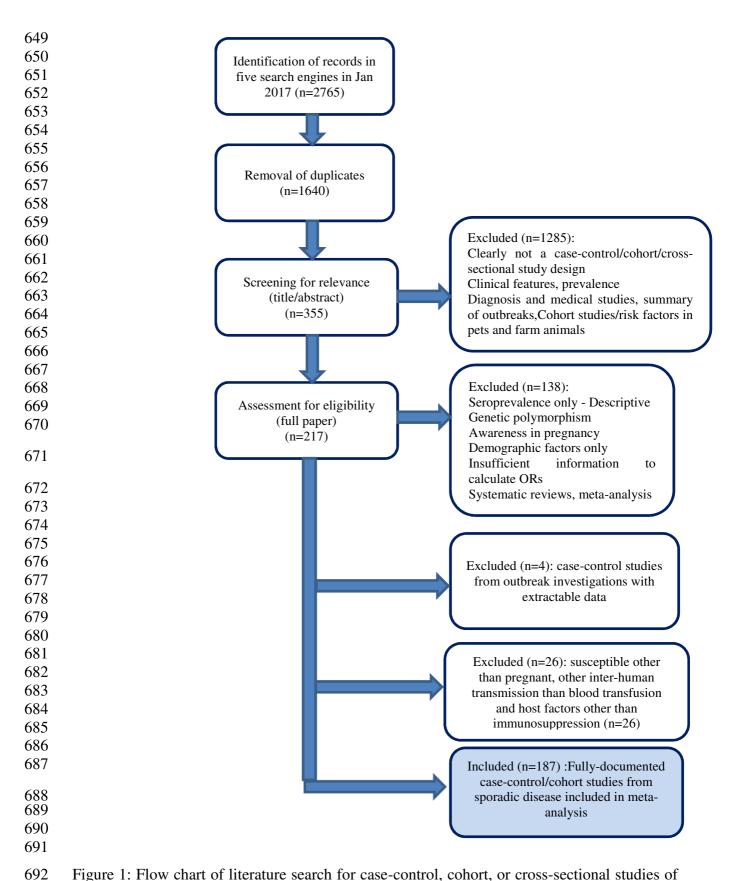


Figure 1: Flow chart of literature search for case-control, cohort, or cross-sectional studies of human sporadic toxoplasmosis

Study	Country	Label	Odds Ratio	[95% CI]	
Alvarado_BMJ_2016	Mexico	Blood transfusion	0.65	[0.04-11.49]	
Alvarado-Esquivel_BMC_2006	Mexico	Blood transfusion	2.37	[0.64-8.68]	.—
Alvarado-Esquivel_BMC_2006*	Mexico	Blood transfusion	28.9	[0-250]	←
Awoke_APJTM_2015	Ethiopia	Blood transfusion	3.68	[0.96-14.06]	-
Awoke_APJTM_2015*	Ethiopia	Blood transfusion	3	[0.74-12.14]	-
Gelaye_IntJInfDis_2015	Ethiopia	Blood transfusion	0.76	[0.16-3.64]	_
Gelaye_IntJInfDis_2015*	Ethiopia	Blood transfusion	0.63	[0.12-3.27]	_
Han_JParasitol_2008	Korea	Blood transfusion	1.57	[0.19-12.82]	-
Almushait_JPD_2012	SaudiArabia	Previous history of blood transfusion	1.16	[0.06-21.35]	<
Almushait_JPD_2012	SaudiArabia	Previous history of blood transfusion	2.13	[0.47-9.61]	-
Random Effect Meta-Analysis	All		1.78	[1.03-3.09]	•

Figure 2: Forest plot of the association of blood transfusion with positive *Toxoplasma* serology in pregnant women. (* adjusted OR as described in Gonzalez-Barron *et al.*, 2019)

Study	Country	Label	Odds Ratio	[95% CI]	
Alvarado_BMC_2015	Mexico	Birds at home	1.11	[0.35-3.51]	
Avelino_BJID_2004	Brazil	Contact with cockroaches	1.72	[1.42-2.08]	•
Avelino_EJOGRB_2003	Brazil	Contact with cockroaches	0.77	[0.43-1.4]	_
Avelino_BJID_2004	Brazil	Contact with flies	1.71	[1.4-2.08]	-
Avelino_EJOGRB_2003	Brazil	Contact with flies	1.28	[0.67-2.43]	_
Avelino_BJID_2004	Brazil	Contact with rats	1.83	[1.39-2.42]	-
Avelino_EJOGRB_2003	Brazil	Contact with rats	1.6	[0.98-2.62]	-
Cavalcante_JP_2006	Brazil	Hunting	0.58	[0.32-1.04]	-
Said_EI_2017*	UK	Hunting	2.12	[0.16-Inf]	
Kolbekova_CMI_2007*	CzechRep	Owning a rabbit	1.47	[1.22-1.77]	-
Filahun_EI_2016	Ethiopia	Presence of feral cats	1.69	[1.08-2.63]	-
Filahun_EI_2016*	Ethiopia	Presence of feral cats	1.63	[0.99-2.7]	-
Ferreira_EpiInf_2014	Brazil	Rats, cockroaches or flies in the home	1.06	[0.64-1.75]	-
Random Effect Meta-Analysis	All		1.53	[1.25-1.88]	•

Figure 3: Forest plot of the association of flies/rodents with positive *Toxoplasma* serology in the mixed population. (* adjusted OR as described in Gonzalez-Barron *et al.*, 2019)

Study	Country	Label	Odds Ratio	[95% CI]	
Alvarado_JP_2011	Mexico	Raw cow milk consumption	0.74	[0.44-1.26]	
Alvarado_OM_2011*	Mexico	Raw milk consumption	7.5	[0.4-144]	37
Alvarado_PV_2012*	Mexico	Raw goat milk consumption	2.77	[0.72-10.57]	
Alvarado-Esquivel_JP_2010-1	Mexico	Raw cow milk consumption	1.01	[0.49-2.09]	-
velino_BJID_2004	Brazil	Unpasteurised goat milk	1.24	[0.97-1.6]	· ·
Avelino_EJOGRB_2003	Brazil	Consumption of raw milk	1.55	[0.79-3.05]	
Elsheikha_PR_2009	Egypt	Drink raw milk/ate kareesh cheese	4.77	[2.45-9.29]	
erreira_EpiInf_2014	Brazil	Drink unpasteurised milk	0.86	[0.53-1.39]	-
Flatt_EurJPublHealth_2013	UK	Drink unpasteurised milk	1.51	[1.12-2.03]	-
flatt_EurJPublHealth_2013	UK	Ate unpasteurised or soft cheeses	0.95	[0.78-1.17]	•
flatt_EurJPublHealth_2013*	UK	Drink unpasteurised milk	1.38	[1.01-1.88]	-
latt_EurJPublHealth_2013*	UK	Ate unpasteurised or soft cheeses	0.87	[0.68-1.13]	-
Garcia_EmInfDis_2003*	Brazil	Ate unpasteurised butter	1.4	[1.1-1.8]	
Gebremedhin_BMC_2013	Ethiopia	Raw milk consumption	1.22	[0.63-3.39]	
Gebremedhin_BMC_2013*	Ethiopia	Raw milk consumption	1.31	[0.59-2.9]	·
ones_CID_2009*	USA	Drink unpasteurised goat's milk	5.09	[1.45-17.8]	
//illar_CienciaRural_2007	Brazil	Consumption of raw milk	1.22	[0.62-2.4]	
//inbaeva_PLOS_2013	Kyrgyzstan	Home made sour cream	1.11	[0.67-1.84]	_
Radon_AJ_2004*	Germany	Consumption of raw farm milk at age 6 yr	2.15	[1.23-3.76]	·
Radon_AJ_2004*	Germany	Consumption of raw farm milk	1.44	[0.56-3.75]	
Radon_AJ_2004*	Germany	Consumption of raw farm milk at age 6 yr	1.8	[0.99-3.28]	
said_EI_2017*	UK	Ate dairy	3.47	[0.26-Inf]	
Said_EI_2017*	UK	Raw milk	0.82	[0.05-14.8] <	-
Said_EI_2017*	UK	Ate dairy	3.86	[0.1-Inf] —	
antos_VetParasit_2009	Brazil	Raw milk consumption	1.9	[0.13-54.47] -	-
ilahun_El_2016	Ethiopia	Boiled and raw milk consumption	1.11	[0.67-1.83]	-
ilahun_El_2016	Ethiopia	Raw milk consumption	1.51	[0.79-2.89]	
Random Effect Meta-Analysis	All		1.56	[1.3-1.88]	•

Figure 4: Forest plot of the association of consumption of dairy products with positive *Toxoplasma* serology in the mixed population. (* adjusted OR as described in Gonzalez-Barron *et al.*, 2019)

Study	Country	Label	Odds Ratio	[95% CI]	
Chiang_PLOSONE_2012	Taiwan	Raw mussels consumption	4	[1.2-12.8]	
Chiang_PLOSONE_2012*	Taiwan	Raw oysters consumption	1.3	[0.8-2.2]	-
Chiang_PLOSONE_2012*	Taiwan	Raw clams consumption	Ť	[0.5-1.9]	-
Chiang_PLOSONE_2012*	Taiwan	Raw mussels consumption	5.3	[1.5-19.1]	_
Chiang_PLOSONE_2014	Taiwan	Raw oysters consumption	1.5	[0.6-3.4]	-
Chiang_PLOSONE_2014	Taiwan	Raw clams consumption	3.6	[1.4-9.3]	-
Chiang_PLOSONE_2014*	Taiwan	Raw clams consumption	3.7	[1.4-9.9]	-
Jones_CID_2009*	USA	Ate raw oysters, clams or mussels	2.22	[1.07-4.61]	-
Said_EI_2017*	UK	Ate shellfish	2.37	[0.48-11.7]	-
Random Effect Meta-Analysis	AII		1.92	[1.4-2.64]	•

Figure 5: Forest plot of the association of consumption of mollusks with positive *Toxoplasma* serology in the mixed population . (* adjusted OR as described in Gonzalez-Barron *et al.*, 2019)

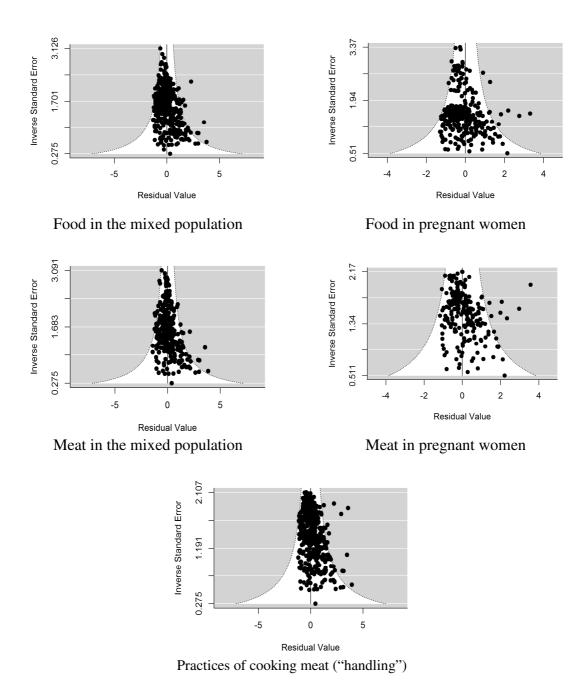


Figure 6: Funnel plots of meta-analyses investigating categorized risk factors (i.e., food in mixed and pregnant population, meat in mixed and pregnant population, and practices of cooking meat)

720 Table 1. Meta-analysis results on main (significant) risk factors for positive *Toxoplasma* serology721

Population	Geographical area	Risk factor	Pooled OR [95% CI]	N/n*	p-val risk factor	Publication bias p-value	Points removed	Heterogeneity analysis***
				Travel	I		1	
Pregnant	All	Abroad	1.878 [1.284 - 2.746]	6/7	0.001	0.754	0	T^2 =0.083 Q(df=6) = 11.08; p-val = 0.086 S^2 =0.380 I^2 =17.94
				t specific				
Mixed	All	Immuno-compromising conditions	2.407 [1.483 - 3.909]	14/32	0.001	0.188	0	T ² =0.175 QE(df=41) = 128.1;p-val < .0001 s ² =0.955 l ² =15.451
Pregnant	All	Blood Transfusion	1.785 [1.031 - 3.089]	6/10	0.039	0.063	0	T ² =0.003 QE(df=23) = 14.51; p-val = 0.911 s ² =0.812 l ² =0.368
				nal Hygiene				
All	All	Poor personal hygiene	2.023 [1.693 - 2.416]	3/9	<.0001	0.743	0	$\tau^{2}=0$ Q(df=8) = 6.596;p-val = 0.581 s ² =0.241 ² =0
	•		A	nimals		•		
Mixed(at)	All	Farm animals Occupational Pets	1.482 [1.099 -1.998] 2.035 [1.641 - 2.522] 1.759 [1.496 - 2.067]	6/8 20/65 67/137	0.009 <.0001 <.0001	0.176	0	τ ² =0.801 QE(df=218)= 838.7; p-val < .0001 s ² =0.555
		flies/rodents	1.534 [1.249 - 1.882]	8/13	<.0001			I ² =59.1
		Occupational	1.557 [1.245 - 1.948]	8/9	0.0001			т²=0.163
Pregnant	All	Pets	1.536 [1.374 - 1.717]	71/176	<.0001	0.214	0	QE(df=198)= 511.4;p-val < .0001
riegnant	All	flies/rodents	1.470 [1.133 - 1.908]	4/6	0.004	0.214	U	s ² =0.553 l ² =22.75
Children	All	Pets Wild	1.634 [1.331 - 2.005] 1.435 [1.052 - 1.959]	14/74 3/6	<.0001 0.023	0.234	0	r^2 =0.539 QE(df=82) = 413.7; p-val < .0001 s^2 =0.356 l^2 =60.27

			Env	rironment				
		Untreated drinking water	1.429 [1.217 - 1.677]	32/64	<.0001	0.061	0	т2=0.588
	Africa removed	Farm environment	1.363 [1.146 - 1.621]	37/62	0.001			QE(df=197)= 709.0;p-val < .0001
Mixed	(22 ORs)	Playground	1.656 [1.403 - 1.953]	41/66	<.0001			s ² =0.359
		Waste water****	1.523 [1.047 - 2.214]	4/6	0.028			l ² = 62.03
		Untreated drinking water	1.487 [1.279 - 1.729]	34/54	<.0001	0.187	0	τ ² =1.071
Dramant	All	Farm environment	1.804 [1.465 - 2.221]	32/52	<.0001			QE(df=189)= 534.4; p-val < .0001
Pregnant	All	Playground	1.462 [1.319 - 1.621]	47/74	<.0001			s ² =0.539
		Waste water	1.863 [1.162 - 2.986]	7/11	0.010			I ² =66.50
	Oceania (1 OR	Untreated drinking water	1.403 [1.215 - 1.620]	2/23	<.0001	0.947	0	τ ² =0.178
Children	excluded) and	Farm environment	2.642 [1.768 - 3.946]	3/7	<.0001			QE(df=47) = 64.15; p-val = 0.045
,	Africa excluded (6 OR excluded)	Waste water	1.802 [1.228 - 2.645]	2/5	0.003			s ² =0.203 l ² =46.87
				Food				
		Dairy	1.563 [1.298 -1.882]	18/27	<.0001	0.031		т ² =2.212
Mixed	Oceania excluded	Meat	1.761 [1.570 - 1.974]	66/287	<.0001		4	QE(df = 396) = 1146, p-val < .0001;
WIIXEU	(1 OR excluded)	Produce	1.872 [1.539 - 2.276]	37/58	<.0001			s ² =0.669
		Seafood	1.702 [1.332 - 2.176]	4/12	<.0001			I ² =76.7569
		Dairy	1.521 [1.116 - 2.073]	28/44	0.008	0.002		т ² =1.044
Pregnant	All	Meat	1.960 [1.472 - 2.610]	65/241	<.0001		0	QE(df=36) = 1368; p-val < .0001
	,	Produce	1.651 [1.267 - 2.151]	34/64	0.001			s ² =0.459 l ² =69.46
			Poo	r handling	Į.		ı	1
Pregnant	All	Poor (no handwashing before eating or cooking,	2.000 [1.598 - 2.504]	19/35	<.0001	0.599	0	τ^2 =0.378 QE(df=45) = 84.05; p-val = 0.001 s^2 =0.489
20 ***/ **		no washing knife)				1 ****		12=43.607

*N/n Number of studies/number of OR;** points removed by sensitivity analysis, all results are given after removing data concerned; ***Between-study variability (τ²), test for residual heterogeneity (QE), variance of residuals (s²), intra-class correlation (l²). **** including non-compliant toilets or lack of toilets in the main dwelling; (at) the analysis type is significant: results are given for multivariate estimates

Table 2. Meta-analysis results on disaggregated risk factors for positive *Toxoplasma* serology

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Main risk factor	Population	Geographical area	Specific class or specific population	Pooled OR [95% CI]	N/n*	Risk factor p-val	Publi- cation bias p-val	Points removed **	Heterogeneity analysis***				
			·	Animals									
Cat	All	All	Mixed	1.711 [1.463 - 2.001]	63/112	<.0001	0.465	0	т²=0.607				
			Pregnant	1.631 [1.445 -1.841]	70/150	<.0001			QE(df=311) = 1191;p-val < .0001				
							Children	1.653 [1.328 - 2.057]	13/52	<.0001			l ² =52.37 s ² =0.552
				Food									
Meat	Mixed	Oceania	Other red meats	1.897 [1.480 - 2.431]	23/48	<.0001	0.037	8	т²= 1.499				
		excluded (1 OR	Pork	2.114 [1.411 - 3.169]	15/23	0.001			QE(df=274)= 668.8;p-val < .0001				
		excluded)	Others	1.734 [1.544 - 1.947]	54/145	<.0001			s ² =0.717				
			Poultry	1.623 [1.147 - 2.297]	11/15	0.006	1		I ² =67.63				
			Processed meat	1.365 [1.075 - 1.733]	13/26	0.011	=						
			Beef	1.635 [1.170 - 2.285]	15/23	0.004	=						
Meat	Pregnant	All	Other red meats	1.822 [1.279 - 2.595]	11/33	0.001	0.002	0	r ² =1.608				
			Others	1.592 [1.354 - 1.871]	60/106	<.0001			QE(df=235)= 996.1;p-val < .0001				
			Poultry	1.514 [1.130 - 2.028]	11/19	0.005			s ² =0.579				
			Processed meat	1.532 [1.201 - 1.953]	9/27	0.001			I ² =73.50				
			Beef	2.052 [1.576 - 2.672]	11/29	<.0001							
Other red meats	Pregnant	All	Lamb meat	1.832 [1.148 - 2.922]	14/4	0.011	0.260	0	$T^2=0.762$ QE(df=19) = 41.75;p-val = 0.002 $S^2=0.754$ $I^2=50.27$				
Other red meats	Mixed	All	Lamb meat	2.404 [1.189 - 4.859]	5/6	0.015	0.096	2	т ² =0.859				
			Boar meat	2.487 [1.814 - 3.409]	5/8	<.0001			QE(df=42) = 66.08;p-val = 0.010				
			Goat meat	1.667 [1.212 - 2.294]	7/8	0.002			s ² =0.633				
			Venison/ram/horse meat	1.566 [1.221 - 2.008]	9/12	0.001			I ² =57.615				
Produce	Mixed	All	Vegetables	1.866 [1.491 - 2.335]	33/52	<.0001	0.879	2	T ² =0.297 QE(df=56) = 198.6;p-val < .0001 s ² =0.522 l ² =36.29				
Produce	Pregnant	All	Vegetables	1.372 [1.198 - 1.571]	34/57	<.0001	0.784	0	T ² =0.145 QE(df=62) = 124.3; p-val < .0001; s ² =0.234 ² =38.32				
Seafood	All	All	Mollusks	1.917 [1.395 - 2.636]	4/9	<.0001	0.087	0	т2=0.019				

								QE(df=11) = 14.95; p-val = 0.185 s ² =0.258 l ² =6.867	
728	*N/n Number of studies/number of OR;** points removed by sensitivity analysis, all results are given after removing data concerned; ***Between-study variability (τ²), test for residual								

*N/n Number of studies/number of OR;** points removed by sensitivity analysis, all results are given after removing data concerned; ***Between-study variability (τ²), test for residual heterogeneity (QE), variance of residuals (s²), intra-class correlation (l²)

Table 3. Effect of poor handling on the pooled association between positive *Toxoplasma* serology and consumption of meat, vegetables and dairy

Risk Factor	Risk factor precise	Pooled OR [95% CI]	N/n*	p-val risk factor	Ratio Poor handling to Base [95% CI]	Points removed**	Publication bias p-val	Heterogeneity analysis***
Meat	Raw Undercooked Base	1.876 [1.289 - 2.729] 1.685 [1.209 - 2.347] 1.523 [1.317 - 1.759]	41/71 80/139 56/283	0.075 0.287 <.0001	1.232 [0.979 - 1.551] 1.107 [0.918 - 1.334]	2	0.0001	r ² = 0.229 QE(df=490) =1534;p-val < .0001; s ² =0.229 l ² =50
Fruit & vegetables	Unwashed Base	1.789 [1.204 - 2.658] 1.202 [0.999 - 1.446]	39/59 14/28	0.001	1.489 [1.206 - 1.839] –	0	0.647	r ² =0.502 QE(df=27) = 355.0; p-val < .0001 s ² =0.394 l ² =56.015
Cheese and milk	Raw Base	1.459 [0.939 - 2.267] 1.019 [0.874 - 1.189]	39/52 10/19	0.015 0.802	1.430 [1.073 - 1.905] –	0	0.108	r ² =0.337 QE(df=69) = 195.5; p-val < .0001 s ² =0.349 l ² =49.128

*N/n Number of studies/number of OR;** points removed by sensitivity analysis, all results are given after removing data concerned; ***Between-study variability (\tau^2), test for residual heterogeneity (QE), variance of residuals (s^2), intra-class correlation (l^2).