

### Assessment of Ovarian Tumor Growth in Wild-Type and Lumican-Deficient Mice: Insights Using Infrared Spectral Imaging, Histopathology, and Immunohistochemistry

Pierre Nizet, Valérie Untereiner, Ganesh D Sockalingum, Isabelle Proult, Christine Terryn, Albin Jeanne, Lise Nannan, Camille Boulagnon-Rombi, Christèle Sellier, Romain Rivet, et al.

#### ▶ To cite this version:

Pierre Nizet, Valérie Untereiner, Ganesh D Sockalingum, Isabelle Proult, Christine Terryn, et al.. Assessment of Ovarian Tumor Growth in Wild-Type and Lumican-Deficient Mice: Insights Using Infrared Spectral Imaging, Histopathology, and Immunohistochemistry. 19th European Conference on the Spectroscopy of Biological Molecules (ECSBM), Aug 2022, REIMS, France. hal-03926632

### HAL Id: hal-03926632 https://hal.univ-reims.fr/hal-03926632

Submitted on 6 Jan 2023

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.











**URCAnim** 



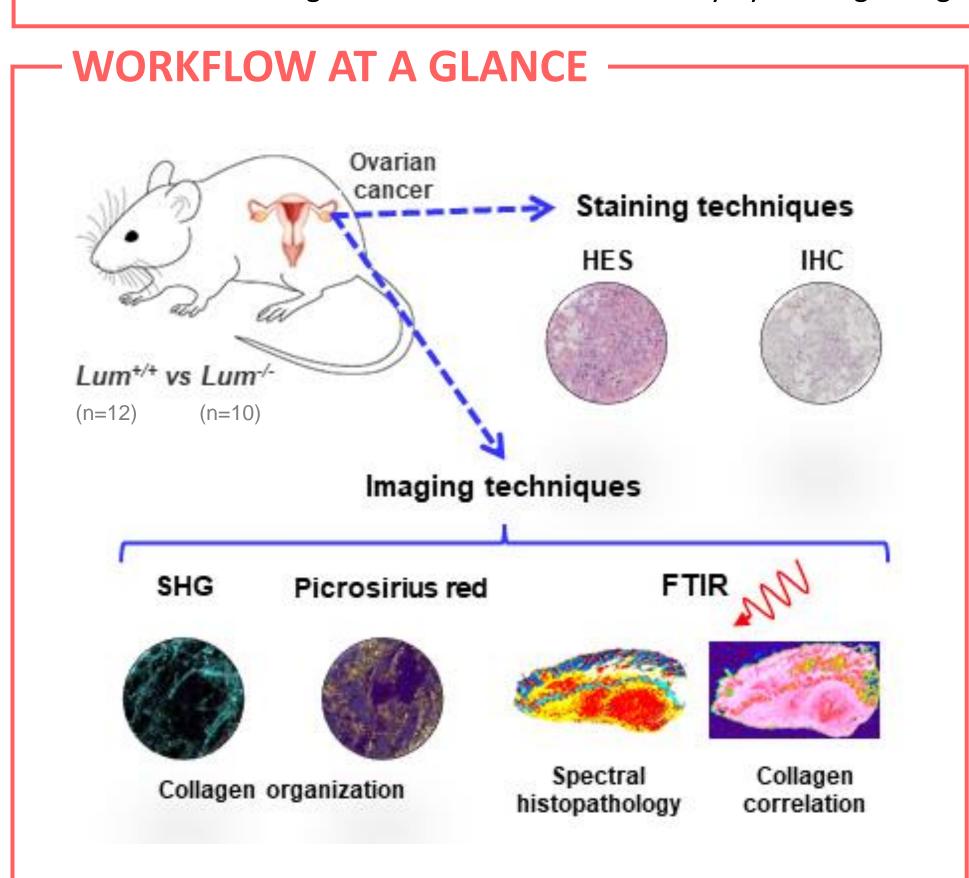
# Assessment of Ovarian Tumor Growth in Wild-Type and Lumican-Deficient Mice: Insights Using Infrared Spectral Imaging, Histopathology, and Immunohistochemistry

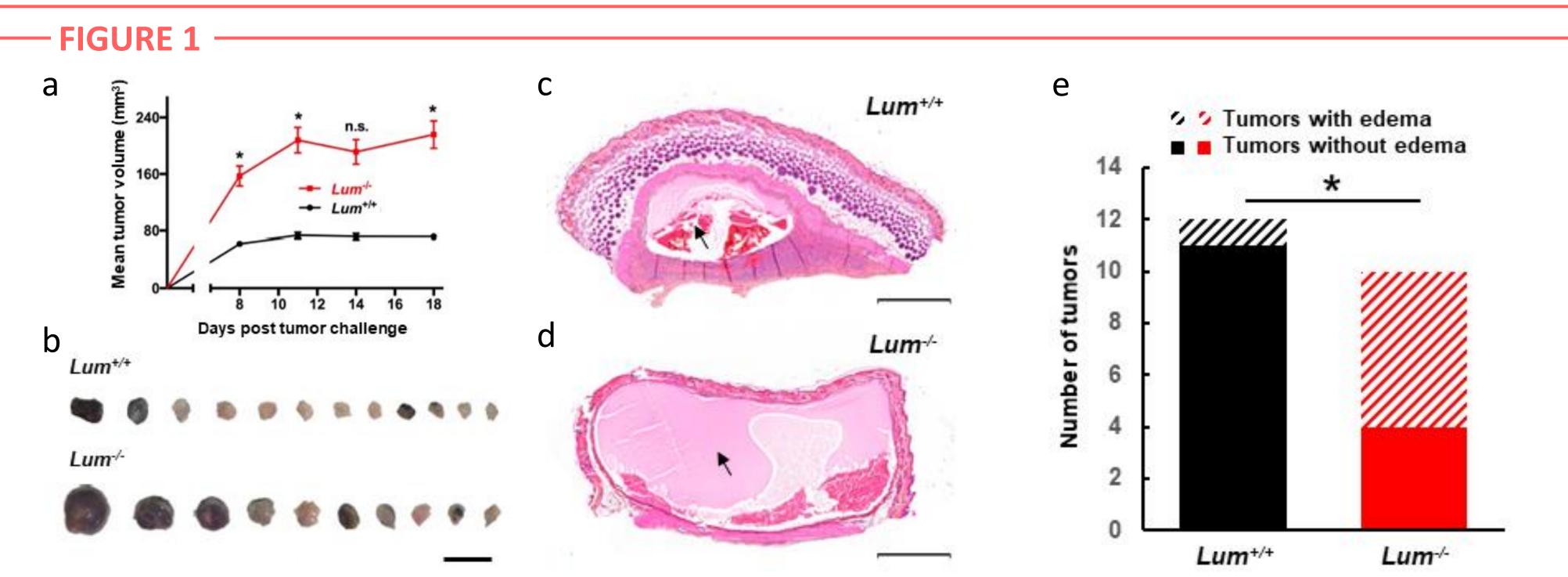
Pierre Nizet<sup>1,2</sup>, Valérie Untereiner<sup>3</sup>, Ganesh D. Sockalingum<sup>4,\*</sup>, Isabelle Proult<sup>1,2</sup>, Christine Terryn<sup>3</sup>, Albin Jeanne<sup>5</sup>, Lise Nannan<sup>1,2</sup>, Camille Boulagnon-Rombi $^{2,6}$ , Christèle Sellier $^{1,2}$ , Romain Rivet $^{1,2}$ , Laurent Ramont $^{1,2,7}$ , and Stéphane Brézillon $^{1,2,*}$ .

<sup>1</sup> Laboratoire de Biochimie Médicale et Biologie Moléculaire, Université de Reims Champagne-Ardenne, 51097 Reims, France; <sup>2</sup> CNRS UMR 7369, Matrice Extracellulaire et Dynamique Cellulaire-MEDyC, 51097 Reims, France; <sup>3</sup> PICT, Université de Reims Champagne-Ardenne, 51097 Reims, France; <sup>4</sup> BioSpecT-EA7506, Université de Reims Champagne-Ardenne, 51097 Reims, France; <sup>5</sup> Apmonia Therapeutics, 51686 Reims, France; <sup>6</sup> CHU Reims, Service de Pathologie, Université de Reims Champagne-Ardenne, 51097 Reims, France; <sup>7</sup> CHU Reims, Service Biochimie-Pharmacologie-Toxicologie, 51097 Reims, France; \* Correspondence: ganesh.sockalingum@univ-reims.fr (G.D.S.), stephane.brezillon@univ-reims.fr (S.B.)

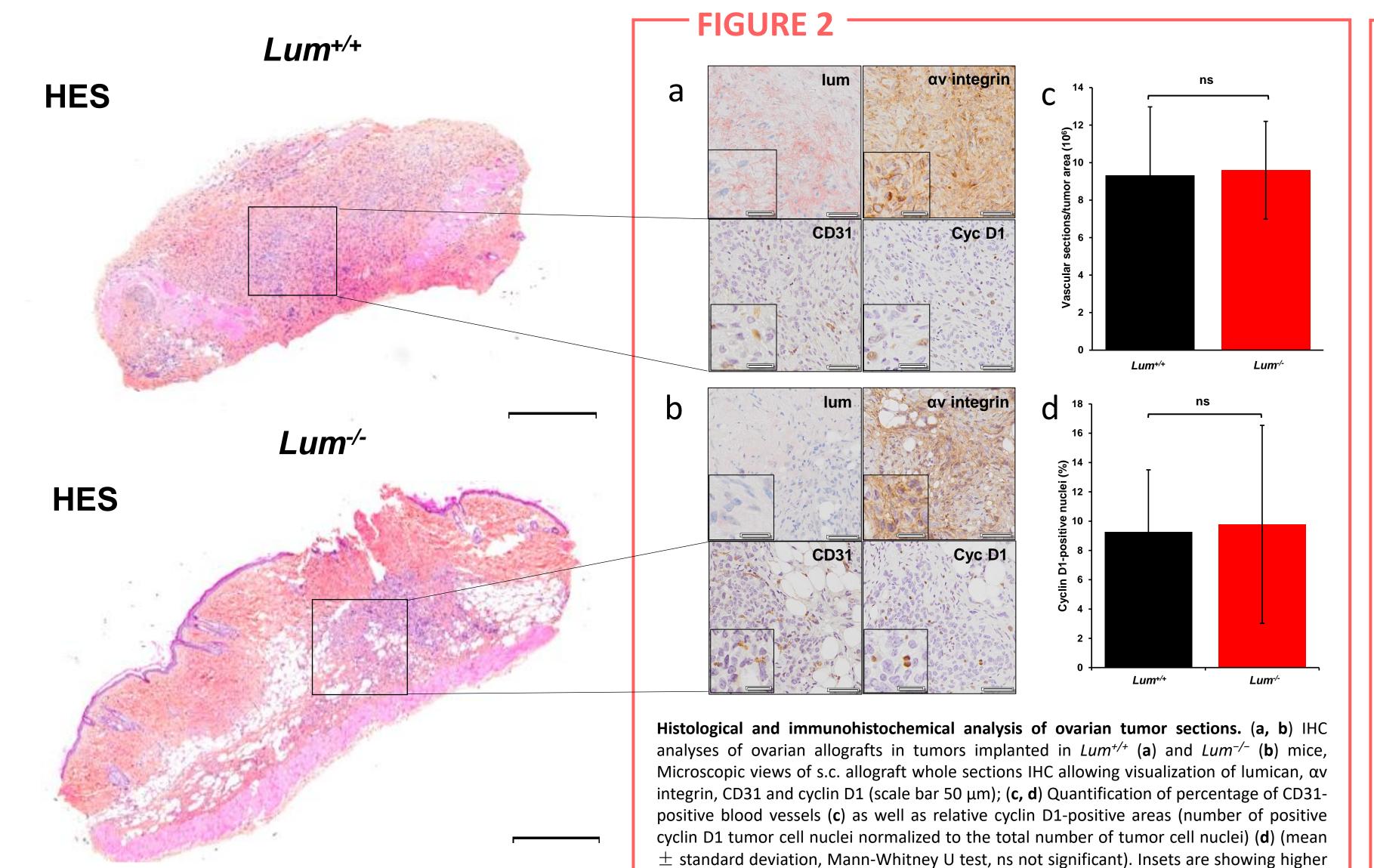
### **ABSTRACT**

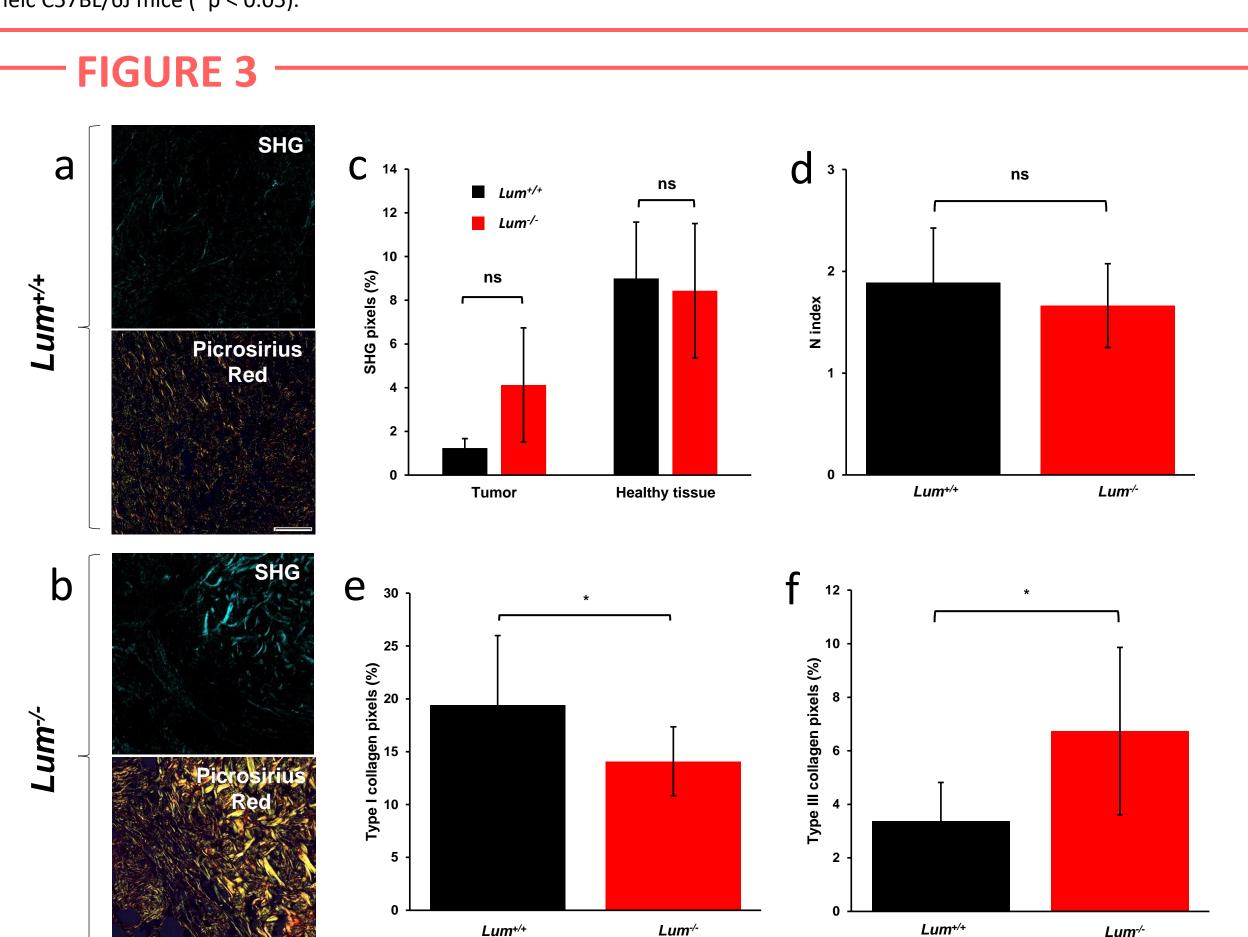
Ovarian cancer remains one of the most fatal cancers because of lack of robust screening methods of detection at early stages. Extracellular matrix (ECM) mediates interactions between cancer cells and their microenvironment via specific molecules. Lumican, a small leucine-rich proteoglycan (SLRP), maintains ECM integrity and inhibits both melanoma primary tumor development and metastatic spreading. The aim of this study was to analyze the effect of lumican on tumor growth of murine ovarian epithelial carcinoma. C57BL/6 wild type mice (n=10) were subcutaneously injected with murine ovarian epithelial carcinoma ID8 cells and sacrificed after 18 days. Analysis of tumor volumes demonstrated an inhibitory effect of endogenous lumican primary tumors were subjected to histological and immunohistochemical staining using antilumican, anti-αv integrin, anti-CD31 and anti-cyclin D1 antibodies, and further examined by label-free infrared spectral imaging (IRSI), second harmonic generation (SHG) and Picrosirius Red staining. The IR tissue images identified different ECM tissue regions of the skin and the ovarian tumor. Moreover, IRSI showed a good correlation of av integrin immunostaining and collagen organization within the tumor. Our results demonstrate for the first time that lumican inhibits the growth of ovarian cancer mainly by altering collagen organization and distribution.



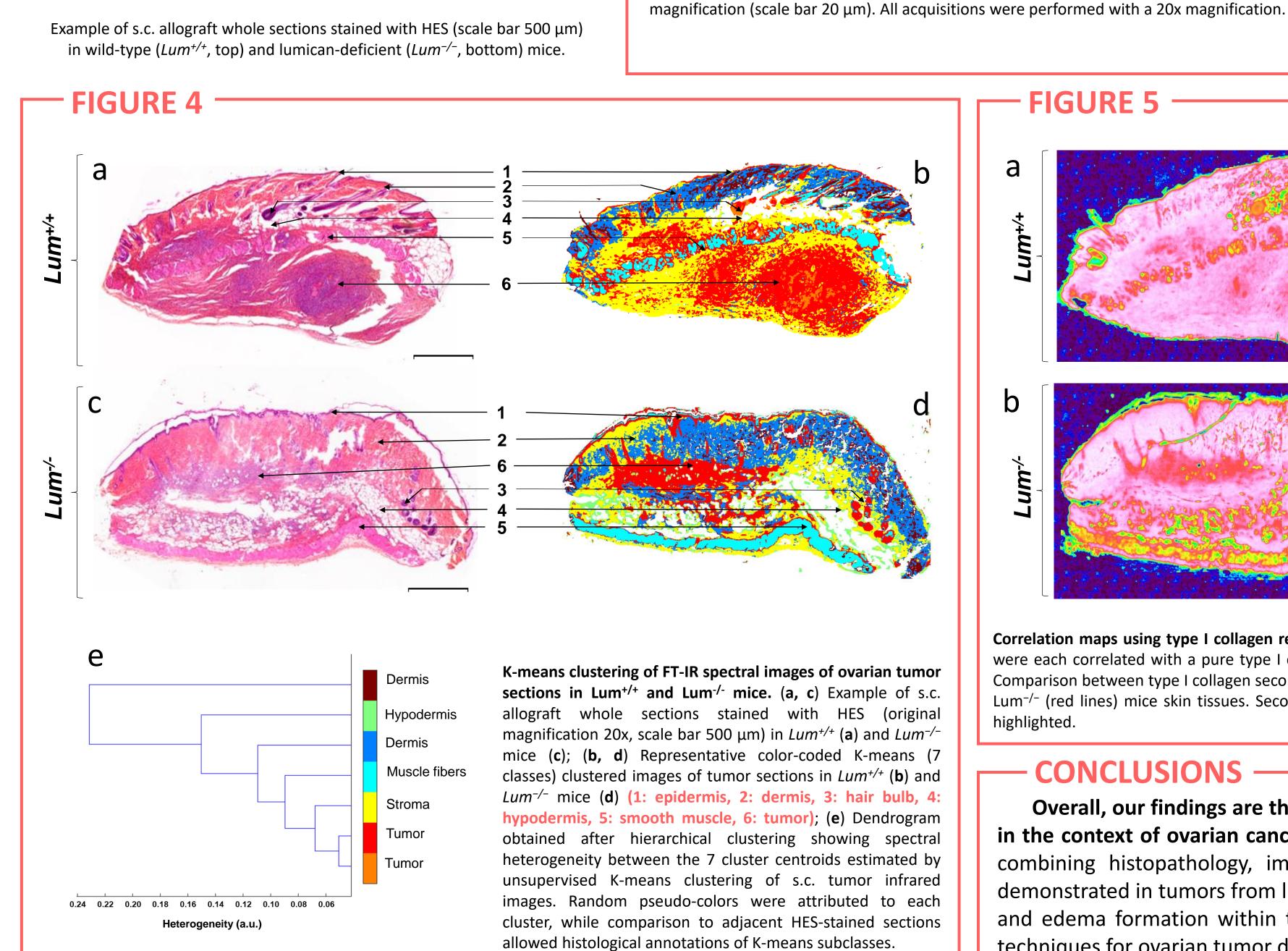


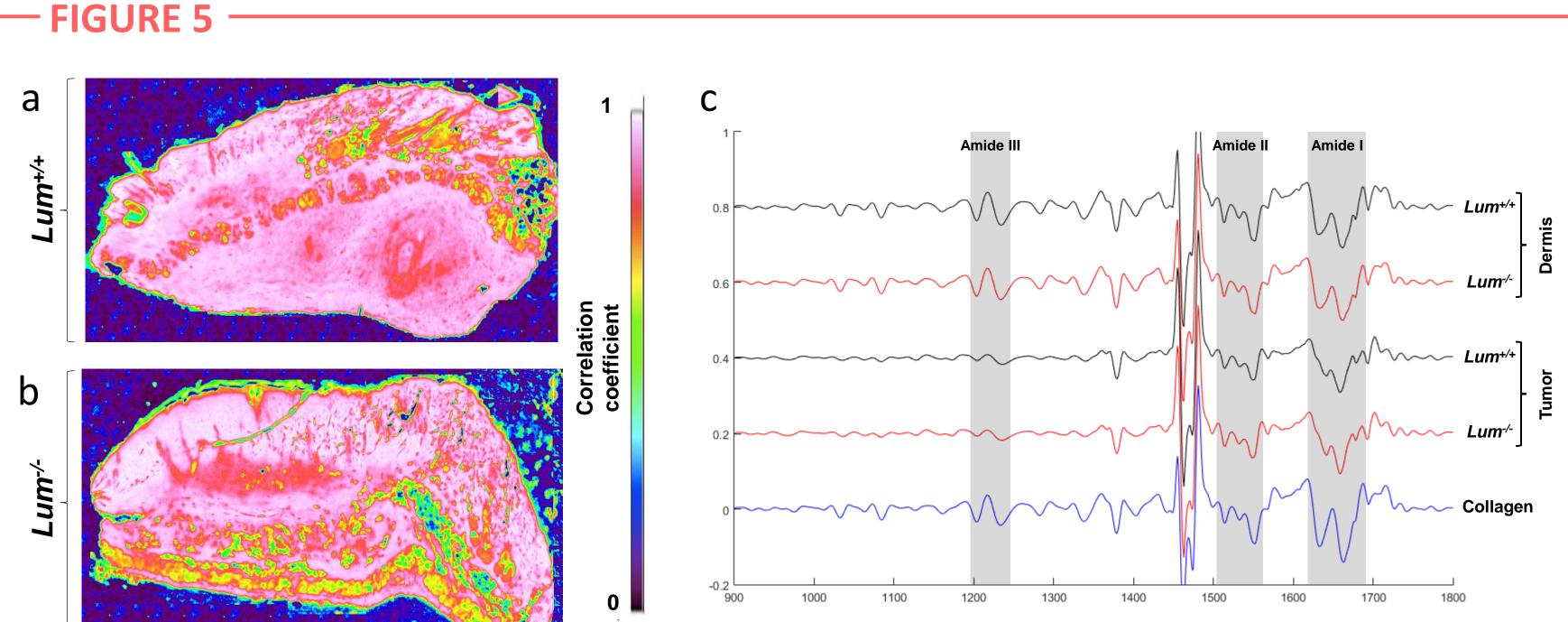
Evaluation of endogenous lumican impact on tumor growth in an ovarian allograft model. (a-b) ID8 ovarian tumor cells (2.5 x 10<sup>5</sup>) were s.c. inoculated in wild-type (Lum<sup>+/+</sup>) or lumican-deficient (Lum<sup>-/-</sup>) syngeneic C57BL/6J mice; (a) Averages of calculated tumor volumes in mm<sup>3</sup> (mean  $\pm$  SEM, n = 10–12 per group) (t test, ns not significant, \*p < 0.05); (b) Representative photographs of ID8 ovarian tumors s.c. allografts after tumor excision (scale bar, 1 cm). Representative images of edemas observed in HES staining of Lum<sup>+/+</sup> (c) and Lum<sup>-/-</sup> (d) tumor sections are shown (scale bar, 500 μm); (e) Quantification of the number of edemas observed in ovarian tumor sections of  $Lum^{+/+}$  or  $Lum^{-/-}$  syngeneic C57BL/6J mice (\*p < 0.05).





Analysis of collagen organization in ovarian tumor sections of wild-type and lumican-deficient mice. (a, b) SHG image analyses and Picrosirius red staining (viewed under widefield cross-polar optics) of ovarian allografts in tumors implanted in  $Lum^{+/+}$  mice (a) and in tumors from  $Lum^{-/-}$  mice (b) (scale bar 50 µm, original magnification 20x); (c) Analysis of collagen fibers intensity by SHG in tumors and healthy tissues present in each section (mean  $\pm$  standard deviation, Mann-Whitney U test, ns not significant); (d) Analysis of tumor ECM collagen organization from images derived from Gabor filtering and FFT, processed on Picrosirius red images (mean  $\pm$  standard deviation, Mann-Whitney U test, ns not significant); (e) Quantification on Picrosirius red stained sections of the relative distribution of red pixels (corresponding to type I collagen) in tumor ECM of Lum<sup>+/+</sup> and Lum<sup>-/-</sup> sections (mean  $\pm$  standard deviation, Mann-Whitney U test, \* p < 0.05); (f) Quantification on Picrosirius red stained sections of the relative distribution of green pixels (corresponding to type III collagen) within tumors of Lum+/+ and Lum<sup>-/-</sup> sections (mean  $\pm$  standard deviation, Mann-Whitney U test, \* p < 0.05).





Correlation maps using type I collagen reference spectrum. (a, b) Type I collagen correlation images of Lum<sup>+/+</sup> (a) and Lum<sup>-/-</sup> (b) spectral images (scale bar 500 μm). The latter were each correlated with a pure type I collagen spectrum. Provided scale indicates the degree of correlation from 0 (black, not correlated) to 1 (white, totally correlated). (c) Comparison between type I collagen second-derivative spectrum (blue line) with second-derivative spectra taken randomly from the dermis and tumors of Lum+/+ (black lines) and Lum<sup>-/-</sup> (red lines) mice skin tissues. Second-derivative spectra are offset for clarity. The corresponding bands of Amide I, II and III, as well as collagen-characteristic zones are highlighted.

## - CONCLUSIONS

Overall, our findings are the first to highlight the major role of lumican in the maintenance of the extracellular matrix integrity in the context of ovarian cancer, showing its inhibitory role in primary ovarian tumor growth. Thanks to a multimodal approach, combining histopathology, immunohistochemistry and imaging techniques, the alteration of collagen organization could be demonstrated in tumors from lumican-deficient mice. This disorganization was associated with a significant increase in tumor growth and edema formation within the tumors. Non-invasive methods such as vibrational spectroscopy might be promising diagnostic techniques for ovarian tumor detection at early stages.